Instructions for use

1. Spin down the contaminated cells at 2000g for 5 min at room temperature. Avoid higher speed as that may collect some of the mycoplasma strains.
2. Add 1 mL PurMa™ MYC-I to 50 ml medium (containing 20 mL FBS) and incubate the contaminated cells in this mixture for 3 days.
3. After 4 days, depending of the source of mycoplasma, PurMa™ MYC-II or PurMa™ MYC-III needs be added to the media.
4. We recommend after the first period of treatment, run two parallel culture plates containing PurMa™ MYC-I followed by PurMa™ MYC- II in the first one and PurMa™ MYC-I and PurMa™ MYC-III in the second culture. See below for more details:
5. At 4th day of treatment with PurMa™ MYC-I, spin down the contaminated cells at 2000g for 5 min and replace it with fresh medium containing 1/200 PurMa™ MYC-I and 1/100 PurMa™ MYC-II (and PurMa™ MYC-III in the second plate).
6. Sub-clone the contaminated cell line in 96 well plates. Dilute the cells to have **ONE SINGLE CELL** in each well. Add 200 uL of fresh medium containing 1/200 PurMa™ MYC-I and 1/100 PurMa™ MYC-II (or PurMa™ MYC-III in the second plate). At this stage cells need extra feeder source. The feeder supplement could be spleen cells (for hybridoma and CHO cells) and excessive amount of FBS (30%) for overexpressed or consistent primary cells.
7. Incubate the cells for 5-7 days without changing the media.
8. At this stage some of the wells become free of mycoplasma and as a result will start to grow and produce a colony.
9. It is imperative to separate the plate of the healthy colonies as soon as they are formed as the failure to do so will cause cross contamination from the contaminated wells.
10. For complete eradication, the above-mentioned procedure needs to be done twice as the mycoplasma could stay inactive inside the cells and gets incorporated in the reproductive system after a period of dormancy.